



Stem Cells and Neurological Diseases

Sumaira Pervaiz, Saba Irshad

Institute of Biochemistry and Biotechnology, University of the Punjab, Lahore, Pakistan

Email: saba.ibb@pu.edu.pk

Rec.Date: Jul 26, 2012 23:42

Accept Date: Sep 10, 2012 23:38

Abstract

Now-a-days many human neurological diseases are not currently curable and result in devastating neurologic sequelae. In the last decade many laboratories are focusing on stem cell treatments for CNS diseases. Out of the many stem cell types that are being tested for neurological treatments, the most common are fetal and adult brain stem cells, embryonic stem cells, induced pluripotent stem cells, and mesenchymal stem cells. So, now Patient-specific iPSC-based modelling of neurogenetic and neurodegenerative diseases is an emerging efficient tool for in vitro modelling to understand CNS diseases and to screen for genes and drugs that modify the disease process. So now-a-days a far more pragmatic approach in the short term might be to use stem cells as chaperones for degenerating nervous tissues, also, the targeted delivery of therapeutic agents could be achieved by modifying stem cells to release specific drugs at the site of transplantation. The exploitation and elucidation of this new 'stem cell pharmacology' has the potential to revolutionise treatment of neurological diseases investigations, and also aimed to replace damaged neurons and gliaby direct transplantation or recruitment of newly generated cells in the adult. Now the next step is to translate these exciting advances from the laboratory into clinically useful therapies.

Keywords: Stem cell technology, CNS, Parkinson's disease, Amyotrophic disease, Alzheimer disease.

Introduction

Most commonly occurring human neurological diseases such as stroke, neurodegenerative disorders, neurotrauma, multiple sclerosis (MS), Parkinson's disease and neuro-developmental disorders are caused by a loss of neurons and glial cells in the brain or spinal cord. These disorders usually cause morbidity and mortality as well as increase social and economic burdens of patients and their caregivers¹. Neural stem cell-based therapies are now being developed to treat a spectrum of neurological conditions once thought to be incurable². This review discusses some of the well-studied neural stem cell types and treatments for neuronal injury and neurological disorders.

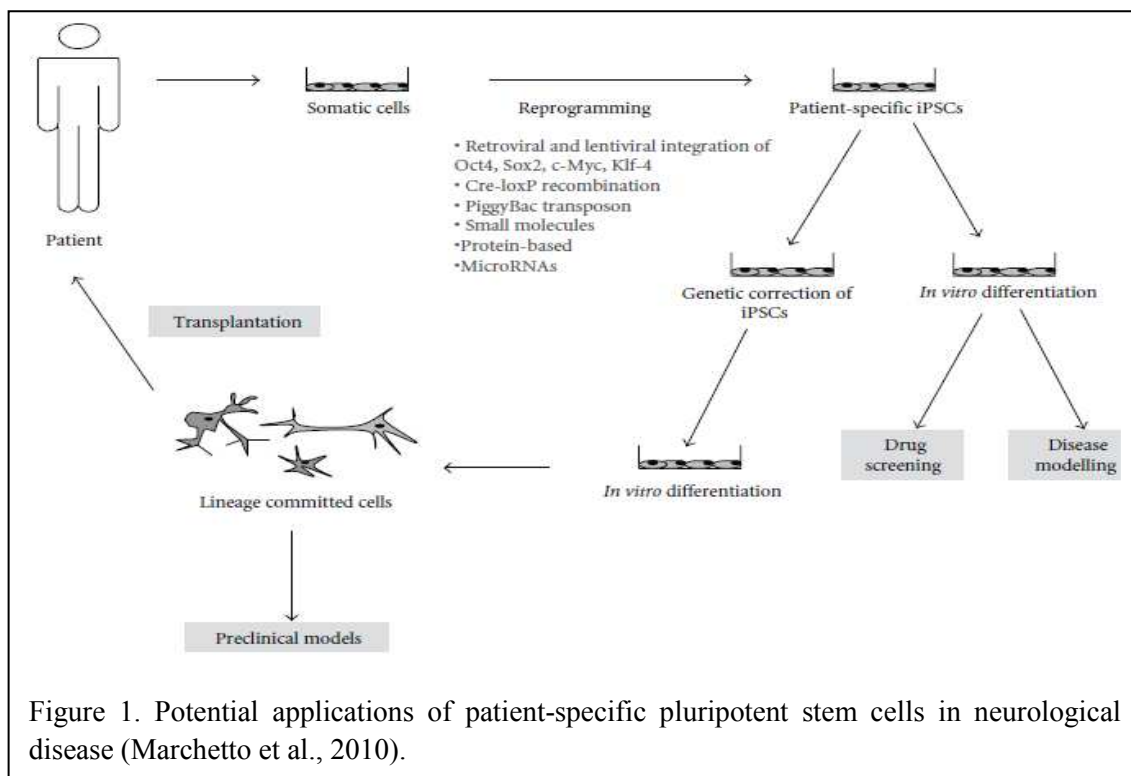
Stem cell plasticity; one cell for all diseases

Stem cells are known by the ability to renew themselves (self-renewal) through mitotic cell division and differentiate into a diverse range of specialized cell types³. They are classified into three types according to their capacity to differentiate into specialized cells (potency). These are totipotent stem cells, pluripotent stem cells (such as embryonic stem cells (ESCs) and induced pluripotent stem cells (iPSCs) artificially derived from a non-pluripotent cell, typically an adult somatic cell through

reprogramming³. ESCs can give rise to all cell types while adult stem cells (ASCs) are thought to be limited in differentiating into different cell types of their tissue of origin. Isolation of ASCs from adult tissue is challenging because they are found in mature tissues however, ESCs can be grown in cell culture⁷². This difference is crucial for stem cell replacement therapies because large numbers of cells are needed for therapeutic applications. The tissues derived from the patient's own ASCs are currently believed less likely to initiate rejection after transplantation. This is significant for solving the immune rejection problem of cell replacement therapies.

The dawn of a new era: induced pluripotent stem cells (iPSCs)

Multipotent stem cells only generate specific lineages of cells. Neural stem cells (NSCs) are multipotent stem cells which are derived from neural tissues⁴. These cells are self-renewing and differentiate into lineage-specific neural precursor or progenitor cells (NPCs) that can give rise to all cell types (neurons, astrocytes, and oligodendrocytes) of the nervous system through asymmetric cell division. So using these stem cells in clinical trials to treat neurological diseases has been started. In this regard, autologous bone marrow stem cells and mesenchymal stem cells are used for treatment of amyotrophic lateral sclerosis. The first clinical trial of an embryonic stem cell-based therapy was authorized in 2009. It was based partly on landmark studies showing functional recovery in rats after spinal cord grafts of oligodendrocyte precursors derived from human embryonic stem cells (hESCs)⁵, the U.S. Food and Drug Administration gave approval to Geron Corporation to begin the first clinical trial of hESC therapy aimed at regenerating myelin in patients with spinal cord lesions⁶. Afterwards, neural stem was approved to test a stem cell therapy in patients with amyotrophic lateral sclerosis.



However, ethical concerns, immune rejection of grafted stem cells, and tumour formation limit the use of human ESCs. So the development of iPSCs in recent years may bypass the ethical controversies and rejection problem using autologous stem cells⁷. In many studies various neural cell types have been differentiated from human or rodent iPSCs generated by the reprogramming of different somatic cells, mainly skin fibroblasts⁸⁻¹⁰. iPSCs have also been differentiated to NPCs^{11, 12}. It has been found that in terms of cell morphology and pluripotency, iPSCs closely resemble ESCs. Several groups have

STEM CELLS AND NEUROLOGICAL DISEASES

successfully generated a wide range of iPSCs from patients with neurodevelopmental and neurodegenerative diseases¹³⁻¹⁵. But before the transplantation process genetic correction of patient-specific iPSCs derived from the patients with neurogenetic disorders is required⁷³. Patient-specific iPSCs also represent a valuable tool to dissect the poorly understood mechanisms of neurogenetic and neurodegenerative diseases⁷³. However the failure to translate the promising results of preclinical neuroprotection studies to the clinic setting may be due to many factors including species differences, brain complexity, age, patient variability, and disease-specific phenotypes that cannot easily be modelled in chosen nonhuman experimental systems^{16, 17}. Cellular modelling studies and chimeric mouse models based on iPSCs may overcome these barriers^{18, 19}. So, the patient-specific iPSCs may be most relevant cell source for drug screening and development as they take into consideration the patient's background, the affected cell type, and the developmental time^{14, 20}.

In this review, we consider several neurological disorders for which stem-cell based therapy has raised particular interest. We describe the ways in which stem cells might be used to treat these conditions, discussing the prospects for and problems of translating laboratory findings into clinically useful therapies.

Parkinson's disease

Parkinson's disease starts at a molecular level (a genetic defect and/or an environmental agent yield to a misfolding of proteins) with consequences at the cellular level (dysfunction of the ubiquitin proteasome system and mitochondria with free radical formation and protein aggregation leading to the apoptotic death of dopaminergic cells), which in turn produces biochemical and neurophysiologic disturbances, particularly dopamine deficiency in the striatum and in other nuclei, and increased firing rate with abnormal firing pattern in the sub thalamic nucleus (STN) and internal segment of the globus pallidus (Gpi). The clinical expression of all these disturbances is Parkinsonism²¹. The main symptoms of the disease are rigidity, poverty of movement (bradykinesia), tremor and postural instability. Current therapies mainly rely on the oral administration of L-dopa and dopamine receptor agonists, and on deep-brain stimulation in the subthalamic nucleus. However, these treatments are effective for some symptoms, but are associated with side effects and do not stop the progression of the disease. So, to be clinically competitive, a stem-cell-based therapy must lead to long-lasting, significant improvement in mobility, ameliorate currently intractable symptoms, or counteract disease progression.

In last twenty years, although the grafting of human fetal tissue-derived dopamine neurons into PD patients has shown some success in small-scale open clinical trials, but this has been tempered by practical limitations such as the use of human fetuses⁷⁵. In efforts to overcome these constraints, expandable sources of stem cells have achieved preliminary success in investigations targeting treatment of PD⁷⁵.

For example, using mouse ES cells, two recent studies have demonstrated the potential utility of these pluripotent populations. Bjorklund et al.,²² showed that undifferentiated naive ES cells, when transplanted into rat models of PD as suspensions of single cells at low concentrations (1 ml of 1000–2000 cells at two striatal sites), spontaneously differentiated into midbrain-like dopaminergic neurons and were able to normalise motor asymmetry in these animals. However, the results were highly variable, with no graft survival in 24% of recipients in addition to frequent teratoma formation that led to death in 20% of transplanted animals. Afterwards, using an improved paradigm, directed differentiation of mouse ES cells into functional dopamine neurons has been achieved with transfection of nuclear receptor related-1 (NURR1 which plays an important role in the proper maintenance of dopaminergic system of brain, mutations can lead to dopamine dysfunction)

STEM CELLS AND NEUROLOGICAL DISEASES

⁷⁴ followed by a multi-step in vitro growth condition-guided approach that takes into account known signals and gene expression patterns during central nervous system (CNS) development ²³. It was found that these differentiated neurons release dopamine, express many key dopaminergic markers and, when grafted into animal models of PD, integrate into the host striatum and significantly improve motor behaviour. Although Kim et al.,²³ enriched differentiated neurons to minimise undifferentiated dividing cells and did not observe teratomas, they cautioned that tumour formation remains a primary concern; thus, the long-term safety and functional benefits of these grafts must be carefully investigated. Moreover, human ES cells might require a different set of signals for dopaminergic maturation and survival in similar models.

Elucidation of the mechanisms underlying the therapeutic effects of stem cell transplantation, it was proposed that, in addition to neuronal replacement, undifferentiated neural stem cells might act as chaperones that offer neuroprotection and mediate rescue of degenerating host populations ⁷⁶. Several studies indicated that in some cases, this has been linked to glial cell line-derived neurotrophic factor (GDNF), a potent growth factor with known ‘dopaminotrophic’ effects ²⁴ for example, Ourednik et al.,²⁴ showed that unilateral grafting of immortalised cerebellar neural precursor cell lines (C17.2) into a bilaterally lesioned mouse model of PD, in which dopamine neurons survive but critical enzymes such as tyrosine hydroxylase (TH) downregulate, resulted in bilateral TH re-expression and functional recovery by two weeks after transplantation. Moreover, the majority of surviving TH-positive neurons in the substantianigrawas host-derived, rather than from the grafted precursors. In support of the study by Lie et al.,²⁵, none of the dopamine neurons incorporated bromodeoxyuridine, showing that they had not arisen from proliferating host stem cells. As most donor cells remained undifferentiated and widely dispersed throughout the mesencephalon, and also expressed GDNF, it was proposed that these immortalised neural stem cells might have the ability to establish suitable trophic environments and allow endogenous repair of dysfunctional dopaminergic neurons.

This finding was consistent with a previous study in which GDNF-secreting C17.2 cells exhibited neuroprotective effects on degenerating dopamine neurons in a different mouse model of PD ²⁶. Although these collective findings demonstrate the potential application of such cells in neurodegenerative diseases of a progressive nature, issues remain concerning the safety of immortalised neural stem cell lines; transformed populations have often resulted in cancers of various types, and tumours can be epigenetically induced by growth conditions ^{27,28}. Afterwards, increased survival of dopamine neurons was similarly observed in a rat model of PD by using non-transformed embryonic neural precursors that produce GDNF through lentivirus-mediated genetic modification ²⁹. Recently, it was found that direct GDNF infusion into the putamen has shown significant clinical improvements with minimal side effects in five PD patients, following one year of chronic administration with programmable pumps ³⁰. So taken together, the use of stem cells in conjunction with growth factor treatment, as well as ongoing development of techniques to introduce foreign genes into cellular substrates ³¹, holds great potential for PD and side by side warrants further investigation.

Multiple Sclerosis

Multiple Sclerosis (MS) is defined as an autoimmune disorder in which aberrant immune responses lead to T-cell mediated focal myelin destruction and secondary oligodendrocyte and axonal damage. The current available disease-modifying therapies in MS are based on the idea that modulation of the autoimmune mechanisms will lead to a reduction of inflammatory infiltrates in the central nervous system (CNS) white matter. So, this reduction, in turn, should result in less demyelination and neurodegeneration and should therefore lead to suppression of clinical

manifestations³². However these therapies are not sufficient for a permanent cure of the disease, so due to the limited effectiveness of the available therapies, the assessment of alternative therapeutic strategies in patients with aggressive clinical course is justified. We now know that bone-marrow transplantation is the standard treatment for several haematological malignant disorders and is being assessed for the treatment of severe forms of many autoimmune diseases including MS³². It was found that in patients, intense immunosuppressant therapy might eradicate the defective immune-system thus allowing the reconstitution of a CNS permissive/tolerant environment by a transplanted healthy hematopoietic stem cell compartment. These cells can be obtained from siblings or an unrelated donor closely matched on HLA (allogeneic transplantation), an identical twin (syngeneic transplantation) or from the patient before chemotherapy (autologous transplantation). The hematopoietic progenitor cells can be directly harvested from the bone marrow or collected from peripheral blood. The well known term hematopoietic-stem-cell transplantation (HSCT) includes both sources. Regarding the mortality rate, allogeneic HSCT is associated with up to 40% mortality if the donor is not a sibling. By contrast, the mortality from autologous HSCT typically is less than 10%³². Now-a-days HSCT has been so widely applied to MS patients and clinical efficacy appears so promising that the use of other sources for stem cells is today clinically insignificant, so this facilitated the procedure avoiding the issue of the embryonic stem cells (ES) and the related ethical concerns³³.

Recent studies indicated that Myelin-producing oligodendrocyte progenitor cells (OPCs) are abundant in the adult human brain. It was found that spontaneous remyelination occurs to varying degrees in the early stages of MS, and OPCs are also present in chronic demyelinated MS lesions. Recently, Back (2005),³⁴ showed that astrocyte derived hyaluronan accumulated in demyelinated lesions from MS patients and prevented the maturation of endogenous OPCs.

Afterwards, the transplantation of remyelinating cells represents another approach for treating myelin loss in MS. In this regard human adult and ES-cell-derived OPCs have been shown to myelinate dysmyelinated mouse brain and spinal cord after transplantation. However, a major obstacle is that the inflammatory environment could destroy the grafted OPCs and inhibit their maturation. So in this case, immunosuppressive and anti-inflammatory treatments might therefore be necessary. Another problem is that the demyelinated MS lesions are distributed across multiple locations throughout the CNS. So an effective therapy will require that these implanted OPCs should migrate to all these sites. Studies in this regard showed that, after systemic administration in mice, NS cells migrated to inflammatory demyelinating lesions, where some became OPCs and remyelinated axons³⁵. However, most cells remained undifferentiated and suppressed proinflammatory mechanisms³⁶.

In order to establish the utility of human derivatives for direct remyelination, recent approaches have included prospective isolation of human progenitor cells from the adult postmortem brain, cell-sorting based on the expression of specific markers, and transplantation into demyelinated animal models. Recently, Windrem et al.,³⁷ reported that human cells could integrate into the host brain and replace lost myelin, although the functional effects of such transplants were not established. Thus, a direct correlation between grafted human cells and behavioural recovery is currently missing.

Stroke

Stroke is known to be caused by blockage of a cerebral artery that leads to focal ischaemia, loss of neurons and glial cells, and motor, sensory or cognitive impairments. It has been demonstrated that endogenous precursors proliferate and migrate in response to ischemic brain injury and to possess latent regenerative potential. Till now, no effective treatment to promote recovery exists, so a therapy

STEM CELLS AND NEUROLOGICAL DISEASES

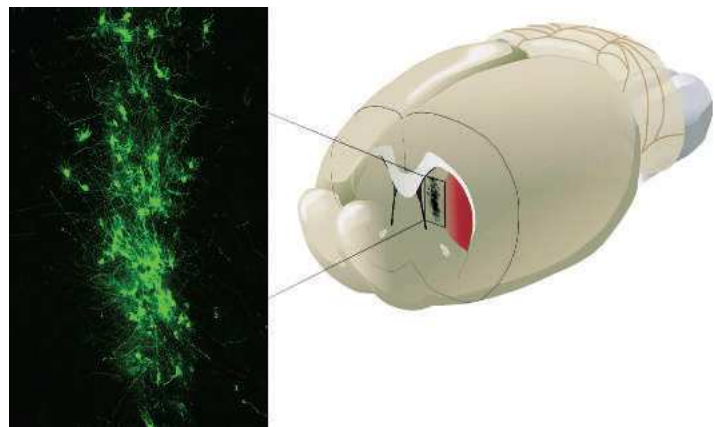
that produced even minor improvement would be valuable. Several studies indicated that transplanted cells from different sources, such as fetal brain, neuroepithelial or teratocarcinoma cell lines, bone marrow and umbilical cord, have yielded some improvement in animals and, in one clinical trial, in humans affected with stroke³⁸.

For enhancing the cell survival and function, mostly the grafts have acted by providing trophic factors³⁸. For stem-cell therapy to be of major clinical value, human cells should be able to replace dead neurons, remyelinate axons and repair damaged neural circuitries. In doing so, human fetal neural stem (NS) cells were transplanted into the brains of stroke-damaged rats, resulting in the migration of new neurons towards the ischaemic lesion³⁹. Similarly, some other studies showed that monkey ES-cell-derived progenitors transplanted into the brains of mice after stroke differentiated into various types of neuron and glial cell, re-established connections with target areas⁴⁰, and led to improved motor function⁴¹. Afterwards it was found that the therapeutic efficacy of such strategies could be improved further by genetically modifying the stem cells: for example, by over expressing an anti-apoptotic gene⁴². Interestingly, the stroke-damaged adult rodent brain has some capacity for neuronal replacement from its own NS cells. For several months after a stroke, NS cells can generate new striatal neurons that migrate to the site of damage^{38, 43}.

A lot of research has been carried out to establish whether endogenous neurogenesis contributes to functional recovery after stroke, and whether this occurs in humans. Also, because the regeneration of cortical neurons will be the basis for functional improvement in most stroke-damaged brains, we will also need to know whether the adult brain's own NS cells can be triggered to produce cortical neurons. So now effective therapies will depend on strategies to increase the survival of the new neurons and to enhance their incorporation into reorganizing neural circuitries.

In this regard, Nakatomi et al.,⁴⁴ reported that, following transient forebrain ischemia which induced selective degeneration of hippocampal CA1 pyramidal neurons, endogenous progenitors from the periventricular region and parenchyma produced new neurons that participated in hippocampal regeneration. It was found that significant recovery levels were noted 28 days post-ischemia, and could be considerably enhanced by treatment with epidermal growth factor and fibroblast growth factor-2. Moreover, regenerated neurons undergo migration to the damaged striatum and express striatal-specific markers. Despite the promise of selfrepair in the adult brain through generation of new neurons, the overall level of endogenous self-recovery in this study was low, i.e. only 0.2% replacement was observed.

Figure 2. Transplantation of stem cells into injured brain. Human fetal NS cells labelled with green fluorescent protein survive for at least 1 month and differentiate into cells morphologically resembling neurons after grafting in close proximity to the stroke-damaged rat striatum (red area) (Lindvall et al., 2006).



Alzheimer's disease

Alzheimer's disease (AD) is an age-related progressive neurodegenerative disorder that is characterized by memory loss and severe cognitive decline. It was found that the neuropathological

STEM CELLS AND NEUROLOGICAL DISEASES

hallmarks of AD are the presence of amyloid β ($A\beta$) peptides in the form of amyloid plaques in the brain parenchyma, particularly in the hippocampus and cerebral cortex, leading to neuronal loss. It has been suggested that $A\beta$ play an etiological, pivotal and likely causal role in the pathogenesis of AD^{45, 46}. Therefore, a reduction of brain $A\beta$ would have the potential to prevent and treat AD⁴⁷.

In AD, patients' memory and cognitive performance is progressively impaired; they develop dementia; and are likely to die prematurely. Current treatment includes the use of acetylcholinesterase inhibitors to enhance cholinergic function, give only partial and temporary alleviation of symptoms. It was found that the pathological changes seen in AD offer an extremely problematic situation for cell replacement. Due to the widespread and progressive damage in the brains of patients with AD, it is unlikely that the mechanisms for instructing transplanted NS cells to differentiate into new neurons will be intact. Theoretically, cognitive decline caused by the degeneration of basal forebrain cholinergic neurons could be prevented by transplanting cholinergic neurons generated from NS cells in vitro. However, to provide long-lasting symptomatic benefit, this approach would require the existence of intact target cells within the patient's brain, and these are highly likely to be damaged. Since stem cells can be genetically modified and have migratory capacity after transplantation, they could be used for the delivery of factors that can modify the course of the disease. So in support of this approach, basal forebrain grafts of fibroblasts that produce nerve growth factor (NGF) which counteracts cholinergic neuronal death, stimulates cell function and improves memory in animal models have been of some benefit in patients with AD⁴⁸.

Recently, the therapeutic potential of bone marrow-derived mesenchymal stem cells (BM-MSCs) has been explored in various pathological conditions of the central nervous system (CNS) such as AD. In a study the feasibility of using the BM-MSCs, as a therapeutic agent for AD has been tested. In order to assess this possibility, an acutely induced AD model induced by injecting amyloid β ($A\beta$) into the dentate gyrus (DG) of hippocampus of C57BL/6 mice was used. It was found that intracerebral transplantation of BM-MSCs into the brain of an induced AD model reduced their $A\beta$ levels when compared to sham-transplanted animals. Side by side the diminution of $A\beta$ deposits was accompanied by the activation of microglia. In addition to these findings it was also found that, the activated microglia was located near the $A\beta$ deposits, and their morphology was changed from ramified to amoeboid as a sign of microglial phagocytosis. This study provides evidence that BM-MSCs can promote the reduction of $A\beta$ through the microglial activation in this acutely induced AD brain, suggesting a potential therapeutic agent against AD⁵⁰.

Huntington's disease

Huntington's disease (HD) is known as a fatal, intractable disorder that is characterized by chorea (excessive spontaneous movements) and progressive dementia. It is caused by the death of projection neurons in the striatum. In this case, stem-cell therapy aims to restore or preserve brain function by replacing and protecting striatal neurons, a strategy that might be insufficient

STEM CELLS AND NEU

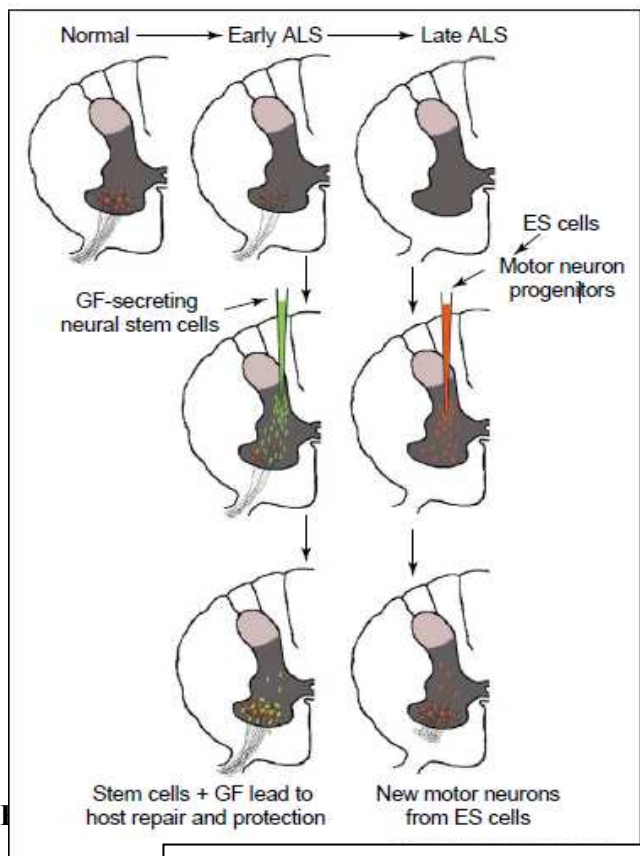


Figure 3. Stem cell- and growth factor (GF)-mediated repair of ALS (Klein et al., 2005)

because patients also suffer progressive neocortical degeneration. Several studies indicated that in animal models of HD, cell replacement using grafts of fetal striatal neurons promotes functional recovery, and some evidence from clinical trials indicates that this can also occur in patients³⁸. By contrast, stem-cell-based approaches are still in their infancy, and the reconstruction of striatal neural circuitry has not been shown in animals. But recently human NS cells implanted into the brains of rats, were found to reduce motor impairments in experimental HD through trophic mechanisms^{50, 51}. So now-a-days, using stem cells for the delivery of trophic factors and neuroprotection to prevent disease progression seems a more achievable clinical goal in HD than neuronal replacement.

Amyotrophic lateral sclerosis

Amyotrophic lateral sclerosis (ALS) is a fatal, progressive neurodegenerative disease that is characterized by motor neuron cell death in the brain and spinal cord accompanied by rapid loss of muscle control and eventual complete paralysis^{52, 53}. In the present day, the only available therapy, riluzole (Rilutek), extends survival only by a matter of months but has shown reliable effects in several clinical trials. It was thought to work through the modulation of glutamate transmission, thereby reducing excitotoxicity⁵⁴, it is also possible that Rilutek increases trophic factor release from astrocytes⁵⁵. Since in amyotrophic lateral sclerosis (ALS), dysfunction and degeneration of motor neurons occur not only in the spinal cord (lower motor neurons) but also in the cerebral cortex and brainstem (upper motor neurons) so a stem-cell therapy must restore or preserve the function of both upper and lower motor neurons, and new neurons must become integrated into existing neural circuitries. So in this regard, recent reports have shown that it is possible to generate lower motor neurons in vitro from stem cells of various sources, including ES cells and those from the fetal CNS^{56, 57}. Afterwards, it was found that mouse ES-cell-derived motor neurons establish functional synapses with muscle fibres in vitro^{58, 59} and extend axons to ventral roots after transplantation into adult rats. But whether these neurons can integrate into existing neural circuitries and restore motor function has not been established. Although neuronal replacement in ALS patients seems a distant goal, using stem cells to prevent motor neurons from dying is a more realistic and shorter-term clinical approach. This prospect is supported by studies showing that human embryonic germ cells delivered into the cerebrospinal fluid of rats with motor neuron injury can migrate into the spinal cord and induce motor recovery, probably through neuroprotection. It was found that the efficacy of this approach could be improved by genetically modifying the stem cells to secrete molecules that promote motor neuron survival. A recent study showed that human cortical progenitors that were engineered to express GDNF survived implantation into the spinal cords of ALS rats and released the neurotrophic factor⁶⁰.

Epilepsy

Epilepsy is defined as a common and diverse set of chronic neurological disorders that is characterized by seizures⁶¹. Some definitions of epilepsy require that seizures be recurrent and unprovoked, but others require only a single seizure combined with brain alterations which increase the chance of future seizures⁶². It was reported that epileptic seizures result from abnormal, excessive or hypersynchronous neuronal activity in the brain. Recently, fetal NSC and ESC-derived neural progenitors have been tested for their ability to integrate and restore function in rodent models of epilepsy. One of the therapeutic goals in epilepsy is to restore the normal balance between excitation and inhibition. So work on fetal neural precursor grafts has shown that they can enhance neuronal inhibition or cause hyperexcitability, depending upon the type of tissue that is used for transplantation and the location of the grafted cells⁶³.

Spinal cord lesions

It is believed that spinal cord injuries interrupt ascending and descending axonal pathways, and cause a loss of neurons and glia, inflammation and demyelination. These lesions lead to a loss of movement, sensation and autonomic control below the site of injury. To date, there is no cure, and the most common current treatment i.e., high-dose methyl-prednisolone, is of questionable value. Now it is evident that the transplantation of stem cells into injured spinal cord can lead to functional benefits^{64, 65}, mainly through trophic factor secretion or the remyelination of spared axons. Recently, a study showed that human NS cells implanted into damaged mouse spinal cord generated new neurons and oligodendrocytes, leading to locomotor recovery⁶⁶. However, it was found that there are risks of side effects unless NS-cell differentiation after transplantation is controlled. For example, astrocytic differentiation and aberrant axonal sprouting after NS-cell implantation into injured rat spinal cord can cause hypersensitivity to stimuli that are not normally painful⁶⁷. So the most realistic short-term clinical goal is to use stem cells for remyelination, which probably occurs to some degree after lesions from endogenous OPCs⁶⁸. In a recent study it was reported after NS-cell implantation into injured spinal cord in rats, there was a good correlation between the number of graft-derived oligodendrocytes, the amount of myelin, and the extent of functional recovery⁶⁷. Similarly, another study reported that transplanted oligodendrocytes from human ES cells could myelinate the injured rodent spinal cord and improves motor function⁶⁹.

Conclusion and future aspects

Stem cells could soon be used to treat neurological disorders, but perhaps not in the way envisioned by most people. The precise mechanisms of stem cell therapy remain to be established, but are probably associated with release of growth factors and other trophic agents into the damaged brain. Future exploitation of stem cell biology, including enhanced release of therapeutic factors through genetic stem cell engineering, might thus constitute promising pharmaceutical approaches to treating diseases of the nervous system.

So firstly, the source of cells must be carefully considered, along with the surgical approach and patient selection. Even if these protective strategies were shown to be safe in patients, they could also pave the way toward improving hES or iPSC-based replacement strategies through increased motor neuron survival and subsequent fiber outgrowth. We can say that the challenges are great, but the rewards are even greater in the continual fight against this devastating disease.

Now-a-days, the iPSC approach to studying brain disease remains one, overall, of tremendous promise. But these are clearly 'early days', and many challenges will need to be overcome before mechanistic insights for major brain diseases, such as Alzheimer's disease, Parkinson's disease, or stroke, are generated via iPSC approaches. In this case perhaps the most critical challenge lies not within the derivation from a given patient and control group of reasonably uniform lines of pluripotent cells in a cost-effective manner, but developing the cell culture and xenograft approaches that will be required to use these lines for the study of brain disease.

In the case of Parkinson's disease, the use of stem cells in conjunction with growth factor treatment, as well as ongoing development of techniques to introduce foreign genes into cellular substrates³¹, holds great potential for PD but also warrants further investigation, similarly in the case of stroke the functional effects of transplants are still not established. In the case of Huntington's disease using stem cells for the delivery of trophic factors and neuroprotection to prevent disease progression seems a more achievable clinical goal as compared to neuronal replacement but still some mechanisms have to be fully understood.

So it would be premature to launch clinical trials to use stem cells to treat neurological disorders. However for each disease, it is now possible to develop a road map that defines the necessary

STEM CELLS AND NEUROLOGICAL DISEASES

scientific and clinical advances required for stem cells to reach the clinic. But before we apply stem-cell therapies to patients, we must be able to control the proliferation and differentiation of stem cells into specific cellular phenotypes and to prevent tumour formation. Furthermore, the efficacy of stem cells and their mechanisms of action should be demonstrated in animal models with pathology and symptomatology resembling the human disease.

Update

Recently a new article has pushed further the idea of neuroprotection by cells within the environment of dying neurons in diseases such as ALS. For example in an elegant study, Clement et al.,⁷⁰ have made chimeric mice that have both wild-type cells and cells expressing the superoxide dismutase-1 mutation. When they studied the pathology of both cell types within the spinal cord, it was clear that wild type cells, probably astrocytes, were able to protect the motor neurons carrying the mutation. This is the major observation that supports the central tenant of the new upcoming idea that the environment surrounding dying neurons is vital to their health and survival.

References

1. Hung CW, Liou YJ, Lu SW. Stem cell-based neuroprotective and neurorestorative strategies. *Int J Mol Sci* 2010; 11: 2039–2055.
2. Kim SU, Vellis J. Stem cell-based cell therapy in neurological diseases: a review. *J Neurosci* 2009; 29: 2183-2200.
3. Leeb C, Jurga M, McGuckin C, et al. Promising new sources for pluripotent stem cells. *Stem Cell Reviews and Reports* 2010; 6:15–26.
4. Conti L, Cattaneo E. Neural stem cell systems: physiological players or in vitro entities. *Nat Rev Neurosci* 2010; 11:176–187.
5. Keirstead HS, Nistor G, Bernal G, et al. Human embryonic stem cell-derived oligodendrocyte progenitor cell transplants remyelinate and restore locomotion after spinal cord injury. *J Neurosci* 2005; 25: 4694-4705.
6. Alper J. Geron gets green light for human trial of ES cell-derived product. *Nat Biotechnol* 2009; 27: 213-214.
7. Yamanaka S, Blau HM. Nuclear reprogramming to a pluripotent state by three approaches. *Nature* 2010; 465: 704–712.
8. Karumbayaram S, Novitsch BG, Patterson M. Directed differentiation of human-induced pluripotent stem cells generates active motor neurons. *Stem Cells* 2009; 27: 806–811.
9. Swistowski A, Peng J, Liu Q. Efficient generation of functional dopaminergic neurons from human induced pluripotent stem cells under defined conditions. *Stem Cells* 2010; 28: 1893–1904.
10. Zeng H, Guo M, Martins K. Specification of region-specific neurons including forebrain glutamatergic neurons from human induced pluripotent stem cells. *PLoS One* 2010; 5: 210-215.
11. Nemati S, Hatami M, Kiani S. Long-term self-renewable feeder-free human induced pluripotent stem cell-derived neural progenitors. *Stem Cells and Development* 2011; 20: 503–514.
12. Sachdeva R, Onsson ME, Nelander J. Tracking differentiating neural progenitors in pluripotent cultures using microRNA-regulated lentiviral vectors. *Nature* 2010; 25:11602–11607.
13. Chamberlain SJ, Li XJ, Lalande M. Induced pluripotent stem (iPS) cells as in vitro models of human neurogenetic disorders. *Neurogenetics* 2008; 9: 227– 235.
14. Marchetto MC, Winner B, Gage FH. Pluripotent stem cells in neurodegenerative and neurodevelopmental diseases. *Human Mole Genet* 2010; 19: 71– 76.
15. Wichterle H, Przedborski S. What can pluripotent stem cells teach us about neurodegenerative diseases. *Nat Neurosci* 2010; 13: 800–804.
16. Gibbons HM, Dragunow M. Adult human brain cell culture for neuroscience research. *Inter J Biochem and Cell Biol* 2010; 42: 844–856.
17. Koch M, Kokaia Z, Lindvall O, et al. Emerging concepts in neural stem cell research: autologous repair and cell-based disease modelling. *The Lancet Neuro* 2009; 8: 819-829.

18. Malgrange B, Borgs L, Grobarczyk B. Using human pluripotent stem cells to untangle neurodegenerative disease mechanisms. *Cellular and Mol Life Sci* 2011; 4: 635–649.
19. Muotri AR. Modeling epilepsy with pluripotent human cells. *Epilepsy and Behaviour* 2009; 14: 81–85.
20. Ebert AD, Svendsen CD. Human stem cells and drug screening: opportunities and challenges. *Nat RevDrug Discovery* 2010; 9: 367–372.
21. Sumaira P, Saba I. Role of nanobiotechnology in Parkinson's disease. *Pak. J. BiochemMolBiol*2012; 45: 35-43.
22. Bjorklund LM, Sanchez PR, Chung S, et al. Embryonic stem cells develop into functional dopaminergic neurons after transplantation in a Parkinson rat model. *ProcNatlAcadSci* 2002; 99: 2344-2349.
23. Kim JH, Auerbach JM, Gomez JA, et al. Neurons derived from embryonic stem cells function in an animal model of Parkinson's disease. *Nature* 2002; 418: 50-56.
24. Ourednik J, Ourednik V, Lynch WP, et al. Neural stem cells display an inherent mechanism for rescuing dysfunctional neurons. *Nat Biotechnol* 2002; 20:1103-1110.
25. Lie DC, Dziewczapolski G, Willhoite AR, et al. The adult substantianigra contains progenitor cells with neurogenic potential. *J Neurosci* 2002; 22: 6639-6649.
26. Akerud P, Canals JM, Snyder EY, et al. Neuroprotection through delivery of glial cell line-derived neurotrophic factor by neural stem cells in a mouse model of Parkinson's disease. *J Neurosci* 2001; 21:8108-8118.
27. Ignatova TN, Kukekov VG, Laywell ED, et al. Human cortical glial tumors contain neural stemlike cells expressing astroglial and neuronal markers in vitro. *Glia* 2002; 39:193-206.
28. Steindler DA, Pincus DW. Stem cells and neurogenesis in the adult human brain. *Lancet* 2002; 359:1047-1054.
29. Ostefeld T, Tai YT, Martin P, et al. Neurospheres modified to produce glial cell line-derived neurotrophic factor increase the survival of transplanted dopamine neurons. *J Neurosci Res* 2002; 69:955-965.
30. Gill SS, Patel NK, Hotton GR, et al. Direct brain infusion of glial cell line-derived neurotrophic factor in Parkinson disease. *Nat Med* 2003; 9:589-595.
31. Pfeifer A, Ikawa M, Dayn M, et al. Transgenesis by lentiviral vectors: lack of gene silencing in mammalian embryonic stem cells and preimplantation embryos. *ProcNatlAcadSci* 2002; 99: 2140-2145.
32. Armitage JO. Bone marrow transplantation. *N Engl J Med* 1994; 330:827–38.
33. Silani V, Cova L, Corbo M, et al. Stem-cell therapy for amyotrophic lateral sclerosis. *Lancet* 2004; 364: 2000–2014.
34. Back SA. Hyaluronan accumulates in demyelinated lesions and inhibits oligodendrocyte progenitor maturation. *Nature Med*2005;11: 966–972.
35. Pluchino S. Injection of adult neurospheres induces recovery in a chronic model of multiple sclerosis. *Nature* 2003; 422: 688–694.
36. Pluchino S. Neurosphere-derived multipotent precursors promote neuroprotection by an immunomodulatory mechanism. *Nature* 2005;436: 266–271.
37. Windrem MS, Roy NS, Wang J, et al. Progenitor cells derived from the adult human subcortical white matter disperse and differentiate as oligodendrocytes within demyelinated lesions of the rat brain. *J Neurosci Res* 2002; 69: 966-975.
38. Lindvall O, Kokaia Z, Serrano A. Stem cell therapy for human neurodegenerative disorders how to make it work. *Nature Med*2004; 10: 23-35.
39. Kelly S. Transplanted human fetal neural stem cells survive, migrate, and differentiate in ischemic rat cerebral cortex. *ProcNatlAcadSci* 2004; 4: 35-36.
40. Hayashi J. Primate embryonic stem cell-derived neuronal progenitors transplanted into ischemic brain. *J Cereb Blood Flow Metab*2006; 4: 45-55.
41. Ikeda R. Transplantation of neural cells derived from retinoic acid-treated cynomolgus monkey embryonic stem cells successfully improved motor function of hemiplegic mice with experimental brain injury. *Neurobiol Dis*2005;20: 38–48.

STEM CELLS AND NEUROLOGICAL DISEASES

42. Wei L. Transplantation of embryonic stem cells overexpressing Bcl-2 promotes functional recovery after transient cerebral ischemia. *Neurobiol Dis*2005; 19: 183-193.
43. Thored P. Persistent production of neurons from adult brain stem cells during recovery after stroke. *Stem Cells*2006; 24: 739–747.
44. Nakatomi H, Kuriu T, Okabe S, et al. Regeneration of hippocampal pyramidal neurons after ischemic brain injury by recruitment of endogenous neural progenitors. *Cell* 2002; 110: 429-441.
45. Hardy J, Selkoe JD. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics, *Science* 2002; 297: 353–356.
46. Selkoe DJ. The molecular pathology of Alzheimer's disease. *Neuron* 1991; 6: 487-498.
47. Wang YJ, Zhou HD, Zhou XF. Clearance of amyloid-beta in Alzheimer's disease: progress, problems and perspectives. *Drug Discov* 2012; 11: 931–938.
48. Tuszynski MH. A phase 1 clinical trial of nerve growth factor gene therapy for Alzheimer disease. *Nature Med*2005; 11: 551–555.
49. Jong KL, Hee KJ, Jae-sung B. Bone marrow-derived mesenchymal stem cells reduce brain amyloid β deposition and accelerate the activation of microglia in an acutely induced Alzheimer's disease mouse model. *Neuroscience*2012; 450: 136–141.
50. Ryu JK. Proactive transplantation of human neural stem cells prevents degeneration of striatal neurons in a rat model of Huntington disease. *Neurobiol Dis*2004; 16: 68-77.
51. McBride JL. Human neural stem cell transplants improve motor function in a rat model of Huntington's disease. *J Comp Neuro*2004;475: 211–219.
52. Cleveland DW, Rothstein JD. From Charcot to Lou Gehrig: deciphering selective motor neuron death in ALS. *Nat Rev Neurosci* 2001; 2: 806–819.
53. Julien JP. Amyotrophic lateral sclerosis: unfolding the toxicity of the misfolded. *Cell* 2001; 104: 581–591
54. Doble A. The pharmacology and mechanism of action of riluzole. *Neurology*1996; 47: 233-241.
55. Peluffo H. Riluzole promotes survival of rat motoneurons *in vitro* by stimulating trophic activity produced by spinal astrocyte monolayers. *Neurosci* 1997; 228: 207–211.
56. Wichterle H, Lieberam I, Porter JA, et al. Directed differentiation of embryonic stem cells into motor neurons. *Cell*2012;4: 385–397.
57. Li XJ. Specification of motoneurons from human embryonic stem cells. *Nature Biotechnol* 2005; 23: 215–221.
58. Harper JM. Axonal growth of embryonic stem cell-derived motoneurons *in vitro* and in motoneuron-injured adult rats. *Proc Natl Acad Sci*2004;101: 7123-7128.
59. Miles GB. Functional properties of motoneurons derived from mouse embryonic stem cells. *J Neurosci*2004; 24: 7848-7858.
60. Klein SM. GDNF delivery using human neural progenitor cells in a rat model of ALS. *Hum Gene Ther*2005; 16: 509-521.
61. Chang BS, Lowenstein DH. Epilepsy. *N Engl J Med*2011; 13: 1257–66.
62. Fisher R, Vanemde BW, Blume W, et al. Epileptic seizures and epilepsy: definitions proposed by the International League Against Epilepsy (ILAE) and the International Bureau for Epilepsy. *Epilepsia*2005; 4: 470–482.
63. Price MG, Yoo JW, Burgess DL, et al. A triplet repeat expansion genetic mouse model of infantile spasms syndrome, Arx(GCG)₁₀ p 7, with interneuronopathy, spasms in infancy, persistent seizures, and adult cognitive and behavioral impairment. *J Neurosci* 2006; 29: 8752-8763.
64. McDonald JW. Transplanted embryonic stem cells survive, differentiate and promote recovery in injured rat spinal cord. *Nature Med*1999; 5: 1410–1412.
65. Ogawa Y. Transplantation of *in vitro*-expanded fetal neural progenitor cells results in neurogenesis and functional recovery after spinal cord contusion injury in adult rats. *J Neurosci Res*2005;69: 925-933.
66. Cummings BJ. Human neural stem cells differentiate and promote locomotor recovery in spinal cord-injured mice. *Proc Natl Acad Sci*2005;102: 14069-14074.
67. Hofstetter CP. Allodynia limits the usefulness of intraspinal neural stem cell grafts; directed differentiation improves outcome. *Nature Neurosci*2005; 8: 346–353.

STEM CELLS AND NEUROLOGICAL DISEASES

68. Yang H. Endogenous neurogenesis replaces oligodendrocytes and astrocytes after primate spinal cord injury. *J Neurosci* 2006; 26: 2157-2166.
69. Keirstead HS. Human embryonic stem cell-derived oligodendrocyte progenitor cell transplants remyelinate and restore locomotion after spinal cord injury. *J Neurosci* 2005; 25: 4694-4705.
70. Clement AM, Nguyen MD, Roberts EA, et al. Wild-type non-neuronal cells extend survival of SOD1 mutant motor neurons in ALS mice. *Science* 2012; 302:113-117.
71. Lindvall O, Kokaia Z. Stem cells for the treatment of neurological disorders. *Nature* 2006; 441: 10-20.
72. Wagers AJ, Weissman IL. Plasticity of adult stem cells. *Cell*; 116: 639-648.
73. Yamanaka S. Strategies and new developments in the generation of patient specific pluripotent stem cells. *Cell* 2009; 1: 39-49.
74. Sacchetti P, Carpentier R, Segard P, Olive-Cren C, Lefebvre P. Multiple signalling pathways regulate the transcriptional activity of the orphan nuclear receptor NURR1". *Nucleic Acids Res* 2006; 34: 5515-27.
75. Mendez I. Dopamine neurons implanted into people with Parkinson's disease survive without pathology for 14 years. *Nat Med* 2008; 14: 38-48.
76. Takagi, Y. Dopaminergic neurons generated from monkey embryonic stem cells function in a Parkinson primate model. *J. Clin. Invest* 2005: 102-109.